This article was downloaded by:

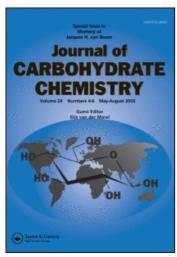
On: 23 January 2011

Access details: Access Details: Free Access

Publisher Taylor & Francis

Informa Ltd Registered in England and Wales Registered Number: 1072954 Registered office: Mortimer House, 37-

41 Mortimer Street, London W1T 3JH, UK



Journal of Carbohydrate Chemistry

Publication details, including instructions for authors and subscription information: http://www.informaworld.com/smpp/title~content=t713617200

An Expeditious Route to *Streptococci* and *Enterococci* Glycolipids *Via* Ring-Opening of 1,2-Anhydrosugars with Protic Acids

C. M. Timmers^a; N. C. R. van Straten^a; G. A. van der Marel^a; J. H. van Boom^a Leiden Institute of Chemistry, Gorlaeus Laboratories, Leiden University, Leiden, The Netherlands

To cite this Article Timmers, C. M., van Straten, N. C. R., van der Marel, G. A. and van Boom, J. H.(1998) 'An Expeditious Route to *Streptococci* and *Enterococci* Glycolipids *Via* Ring-Opening of 1,2-Anhydrosugars with Protic Acids', Journal of Carbohydrate Chemistry, 17: 3, 471 - 487

To link to this Article: DOI: 10.1080/07328309808002906 URL: http://dx.doi.org/10.1080/07328309808002906

PLEASE SCROLL DOWN FOR ARTICLE

Full terms and conditions of use: http://www.informaworld.com/terms-and-conditions-of-access.pdf

This article may be used for research, teaching and private study purposes. Any substantial or systematic reproduction, re-distribution, re-selling, loan or sub-licensing, systematic supply or distribution in any form to anyone is expressly forbidden.

The publisher does not give any warranty express or implied or make any representation that the contents will be complete or accurate or up to date. The accuracy of any instructions, formulae and drug doses should be independently verified with primary sources. The publisher shall not be liable for any loss, actions, claims, proceedings, demand or costs or damages whatsoever or howsoever caused arising directly or indirectly in connection with or arising out of the use of this material.

AN EXPEDITIOUS ROUTE TO STREPTOCOCCI AND ENTEROCOCCI GLYCOLIPIDS VIA RING-OPENING OF 1,2-ANHYDROSUGARS WITH PROTIC ACIDS

C.M. Timmers, N.C.R. van Straten, G.A. van der Marel and J.H. van Boom*

Leiden Institute of Chemistry, Gorlaeus Laboratories, Leiden University, P.O. Box 9502, 2300 RA Leiden, The Netherlands

Received April 1,1997 - Final Form January 5, 1998

ABSTRACT

1,2-Anhydroglucose 6 reacts smoothly and with a high degree of stereoselectivity with a variety of carboxylic and phosphoric acids resulting in the formation of the predominantly β -oriented 1-O-acyl and 1-O-phosphorylglucoses 7-17. This methodology has been successfully applied in the construction of glycolipids 1a,b. Ring-opening of the 1,2-anhydroglucose derivative 19 with benzoic acid furnished exclusively the β -aligned key intermediate 20. Subsequent ICDT-assisted chemoselective α -glucosylation of 20 with thioethyl donor 21, followed by glycosidation of kojibiosyl benzoate 22 with glycerol acceptor 23 gave the fully protected α -diglucosyl glycerol derivative 25, which upon desilylation (\rightarrow 28), acylation (\rightarrow 29 or 30) and deprotection afforded the target glycolipids 1a-b in high overall yield.

INTRODUCTION

It is well documented¹ that glycolipids play a pivotal role as membrane anchors of outer cell-wall components in a variety of organisms. A few years ago, we reported² the

OR¹
OR¹
OR¹
OR¹
OR²
OR²
OR
OP
OP
OP
OP
OP
OP
OR
A

1a
$$R^1 = R^2 = \text{palmitoyl}$$
b $R^1 = R^2 = \text{myristoyl}$

Figure 1

assembly of glycolipid 1 (see Fig. 1, R^1 = stearoyl, R^2 = palmitoyl), which serves as the common metabolic precursor for various $Streptococci^3$ and $Enterococci^4$ glyco(phospho)lipids. Recently, it has been postulated⁵ that $\mathbf{1a}$ (R^1 = R^2 = palmitoyl) and $\mathbf{1b}$ (R^1 = R^2 = myristoyl) may enhance HIV replication. The renewed interest in compound 1 was a stimulus to develop a more straightforward and flexible methodology for the introduction of the requisite α -linkages.

Retrosynthetic analysis (see Fig. 1) reveals that 1 is accessible from a glucosyl donor (2), bearing an appropriate β -oriented anomeric leaving group (Y), a free hydroxyl at position two and a selectively removable 6-O-protecting group (R¹). Chemoselective α -glucosylation of the 2-OH group in 2 with donor 3, followed by coupling of the resulting dimer *via* activation of Y with the suitably protected glycerol acceptor 4, would lead to the α -diglucosyl glycerol core unit of 1.

It was envisaged that the glucosyl donor 2 can be synthesized *via* nucleophilic ring-opening of an appropriately protected α -1,2-anhydroglucose precursor (5), which can be prepared⁶ from the corresponding glucal by 3,3-dimethyldioxirane (DMD)-mediated epoxidation. In a recent paper, Danishefsky *et al.* showed⁷ that 1,2-anhydrosugars can be converted in moderate yields into other glycosyl donors (*i.e.*, 2, Y = SPh, SePh, 4-pentenyl, F) by nucleophilic displacement at the anomeric center. We here report an alternative approach towards 1 based on the ring-opening of the suitably protected α -1,2-anhydroglucose derivative 19 (see Scheme 2) with benzoic acid.

	α:β	yield (%)
7 $R^1 = Ac, R^2 = H$	1:5	87
8 R ¹ = Bz, R ² = H \longrightarrow ii	1:11	91
9 R ¹ = Bz, R ² = Bz ←	1:11	88
10 $R^1 = C(O)H, R^2 = H$ 11 $R^1 = C(O)H, R^2 = Bz$	1:5	72
12 R ¹ = BnO BnO OMe	3:10	70
13 $R^1 = P(O)(OBn)_2$, $R^2 = H$	0:1	82
14 R' = P(O)(OBn) ₂ , R ² = Bz \checkmark	0:1	72
15 R ¹ = P(O)(OBu) ₂ , R ² = H $\frac{1}{100}$	0:1	88
16 R ¹ = P(O)(OBu) ₂ , R ² = Bz \rightarrow	0:1	81
17 R ¹ = BnO BnO OMe	1:4	74

Scheme 1

RESULTS AND DISCUSSION

Prior to the assembly of target glycolipid 1, we investigated the ring-opening of the α -1,2-anhydro function in the known fully benzylated glucopyranose derivative 6^6 with several carboxylic and phosphoric acids (see Scheme 1). Reaction of 6 with a slight excess of acetic acid in anhydrous dichloromethane at 0 °C, proceeded smoothly to afford anomeric acetate 7 in 87% yield with a high degree of stereoselectivity (α : β = 1:5). In a similar fashion, subjection of oxirane 6 to benzoic acid gave the 1-O-benzoyl glucose 8 (91%, α : β = 1:11), one-pot benzoylation of which furnished the dibenzoate 9 (88%). It is

of interest to note that the conversion of 1,2-epoxide 6 with benzoic acid in the more polar solvent THF resulted in a less favorable anomeric mixture of 8 (α : β = 1:3). The latter observation may be attributed to partial dissociation of the acid, inducing ring-opening of 1,2-anhydro derivative 6 into an intermediate oxycarbenium species, which may undergo nucleophilic substitution by benzoate anion from either the α - or the β -side. Treatment of 6 with formic acid in dichloromethane led to the unstable anomeric formate 10 which after benzoylation provided the more stable derivative 11. Apart from this, the ring-opening of oxirane 6 with a more functionalized carboxylic acid derivative was explored. It was established that reaction of 6 with methyl 2,3,4-tri-O-benzyl- α -D-glucuronopyranoside⁸ gave the acyl-linked dimer 12 (α : β = 3:10). The latter type of acyl disaccharides has recently been used in a redox glycosidation strategy.

Interestingly, substitution of 6 with dibenzyl or dibutyl phosphoric acid yielded exclusively the respective β -oriented glucosyl phosphates 13 and 15, benzoylation of which gave the fully protected glucosides 14 and 16.¹⁰ On the other hand, the phosphotriester-bridged diglucoside 17 was obtained as an anomeric mixture (α : β = 1:4) by treatment of 6 with methyl 2,3,4-tri-O-benzyl-6-(benzyl phosphate)- α -D-glucopyranoside.¹¹

On the basis of the ring-opening of 1,2-anhydroglucose 6 with benzoic acid, it was anticipated that treatment of 1,2-oxirane 19, obtained by benzylation of known 6-*O-tert*-butyldiphenylsilyl-D-glucal¹² and subsequent DMD-mediated epoxidation of fully protected glucal 18, with benzoic acid would lead to an anomeric mixture of the corresponding 1-*O*-benzoyl-3,4-di-*O*-benzyl-6-*O-tert*-butyldiphenylsilyl-D-glucopyranose (20). However, it was very gratifying to establish that the nucleophilic displacement proceeded with complete inversion of configuration at the anomeric center to give exclusively the β -aligned key intermediate 20 in 92% yield. Subsequent glucosylation of the 2-hydroxyl in 20 with the known ethyl thioglucosyl donor 21¹³ under the agency of iodonium di-sym-collidine triflate (IDCT)¹⁴ proceeded stereoselectively to give the α -linked disaccharide 22.

At this stage, the kojibiosyl donor 22 was coupled under the agency of trimethylsilyl triflate (TMSOTf)¹⁵ with 1,2-di-*O-tert*-butyldiphenylsilyl-sn-glycerol acceptor 23, readily accessible from commercially available 1,2-O-isopropylidene-sn-glycerol, to give the diglucosyl glycerol derivative 25 as an inseparable anomeric mixture (α : β = 8:1) in 81% yield. Desilylation of 25 with tetra-n-butylammonium fluoride (TBAF) and ensuing separation of the anomeric mixture led to the triol 28. Finally, acylation of 28 with palmitoyl or myristoyl chloride in pyridine/CH₂Cl₂ gave the fully protected glycolipids 29 and 30, respectively, in near quantitative yields. The benzyl

Reagents and conditions: (i) 3,3-dimethyldioxirane, CH_2Cl_2 /acetone, 5 min, quant.; (ii) BzOH, CH_2Cl_2 , 0 °C, 5 min, 92%; (iii) IDCT, $Et_2O/ClCH_2CH_2Cl$ (1:1, v/v), 1 h, 72%; (iv) TMSOTf, $ClCH_2CH_2Cl$, 3 h, 25: 81%, 26: 74%; (v) TBAF, THF, 2-4 h, 27: 83%, 28: 78%; (vi) palmitoyl or myristoyl chloride, pyridine/ CH_2Cl_2 (1:1, v/v), 3 h, 29: 92%, 30: 90%; (vii) H_2 (3 atm.), Pd/C, MeOH/ CH_2Cl_2 (3:1, v/v), 12 h, 1a: 96%, 1b: 94%.

Scheme 2

protective groups in **29** and **30** were removed by hydrogenolysis over Pd/C to afford the respective target molecules **1a** and **1b**, the structure of which was unambiguously ascertained by mass spectrometry and ${}^{1}H$ and ${}^{13}C$ NMR analysis. In addition, TMSOTf-mediated glycosylation of the known 1,2-di-O-allyl-sn-glycerol acceptor **24**¹⁶ with kojibiosyl benzoate **22** led to an inseparable anomeric mixture of the fully protected diglucosyl glycerol derivative **26** (α : β = 6:1). Desilylation of **26** with TBAF followed by separation of the individual anomers provided the partially protected α -diglucosyl

glycerol derivative **27**, which is a useful building block in the construction of various naturally occurring *Streptococci* and *Enterococci* glyco(phospho)lipids.

CONCLUSION

The results presented in this paper show that anomeric acyl or phosphoryl donors are readily accessible by protic acid-mediated ring-opening of 1,2-anhydrosugars. This transformation proceeds predominantly with inversion of configuration at the anomeric center. Furthermore, the high-yielding and efficient synthesis of glycolipids 1a-b may open the way to the preparation of other biologically interesting glyco(phospho)lipids.

EXPERIMENTAL

Materials and methods. ¹H NMR, ¹³C NMR and ³¹P NMR spectra were recorded with a Jeol JNM-FX-200 (200/50.1/80.7 MHz), a Bruker WM-300 (300/75.1/121.0 MHz) or a Bruker DMX-600 spectrometer (600/150.3/242.1 MHz). ¹H and 13 C chemical shifts are given in ppm (δ) relative to tetramethylsilane as internal standard and ³¹P chemical shifts are given in ppm (δ) relative to 85% H₃PO₄ as external standard. Mass spectra were recorded with a Finnigan MAT TSQ70 triple quadropole mass spectrometer. Optical rotations were determined with a Propol automatic polarimeter. Dichloromethane (CH₂Cl₂), diethyl ether, pyridine and toluene were boiled under reflux with CaH₂ for 3 h, distilled and stored over molecular sieves (4 Å). 1,2-Dichloroethane (Biosolvent, HPLC-grade), N,N-dimethylformamide (DMF, Baker, p.a.) and tetrahydrofuran (THF, Biosolvent, HPLC-grade) were stored over molecular sieves (4 Å). Methanol (Rathburn, HPLC-grade) was stored over molecular sieves (3 Å). All other chemicals were obtained from commercial sources and were used as received. Column chromatography was performed on Baker silica gel (0.063-0.200 mm). Gel permeation chromatography was accomplished with LH-20 column material (Sephadex). TLC analysis was done on DC-fertigfolien (Schleicher & Schüll F1500, LS254) with detection by UV-absorption (254 nm) where applicable and charring with 20% H₂SO₄ in MeOH or ammonium molybdate (25 g/L) and ceric ammonium sulfate (10 g/L) in 10% aq. H₂SO₄. Reactions were carried out at ambient temperature, unless otherwise stated. Prior to reactions that require anhydrous conditions, traces of water in the glycosides were removed by coevaporation with 1,2-dichloroethane, pyridine or toluene.

General procedure for the protic acid-mediated ring-opening of 1,2-anhydrosugar 6. To a stirred and cooled solution of 1,2-anhydrosugar 6 (0.43 g, 1.0 mmol) in CH₂Cl₂ (5 mL) was added dropwise, over a period of 5 min, a solution of the

carboxylic or phosphoric acid derivative (1.2 mmol) in $\mathrm{CH_2Cl_2}$ (10 mL).* The reaction mixture was subsequently diluted with $\mathrm{CH_2Cl_2}$ (50 mL) and washed with aq. NaHCO₃ (2 x 25 mL). The organic layer was dried (MgSO₄) and concentrated under reduced pressure. The residue was purified by flash chromatography (10-50% EtOAc/light petroleum) to give an anomeric mixture of the corresponding 1-O-acyl or 1-O-phosphoryl sugar.

*General procedure for the one-pot 2-OH benzoylation. After addition of the appropriate acid to the 1,2-anhydrosugar, pyridine (5 mL) and benzoyl chloride (0.17 mL, 1.5 mmol) were sequentially added and the reaction mixture was allowed to stir for 1 h at room temperature. The reaction mixture was subsequently diluted with CH₂Cl₂ (50 mL) and washed with aq. NaHCO₃ (2 x 25 mL). The organic layer was dried (MgSO₄) and concentrated under reduced pressure. Traces of pyridine in the residue were removed by coevaporation with toluene (3 x 10 mL). Further purification of the residue was accomplished by silica gel chromatography (10-30% EtOAc/light petroleum) to give an anomeric mixture of the corresponding 2-O-benzoyl sugar.

1-*O*-Acetyl-3,4,6-tri-*O*-benzyl-α/β-D-glucopyranose (7). Yield: 0.37 g, 0.87 mmol, 87%, α:β = 1:5. α-anomer: 13 C NMR (CDCl₃): δ 169.5 (C=O Ac), 138.2, 137.7, 137.6 (C_q Bn), 128.2-127.6 (C_{arom}), 91.9 (C₁, J_{C,H} = 170.0 Hz), 82.0, 79.0, 73.4, 71.2 (C₂/C₃/C₄/C₅), 74.8, 74.5, 71.6 (CH₂ Bn), 67.9 (C₆), 20.8 (CH₃ Ac); β-anomer: δ 169.5 (C=O Ac), 138.2, 137.7, 137.6 (C_q Bn), 128.2-127.6 (C_{arom}), 93.8 (C₁, J_{C,H} = 159.4 Hz), 84.4, 76.8, 75.3, 72.8 (C₂/C₃/C₄/C₅), 75.0, 74.5, 73.2 (CH₂ Bn), 67.9 (C₆), 20.8 (CH₃ Ac). MS (ESI): m/z 493 (M+H)⁺, 515 (M+Na)⁺.

1-*O*-Benzoyl-3,4,6-tri-*O*-benzyl-α/β-D-glucopyranose (8). Yield: 0.50 g, 0.91 mmol, 91%, α:β = 1:11. α-anomer: 13 C NMR (CDCl₃): δ 164.8 (C=O Bz), 138.1, 137.9, 137.7 (C_q Bn), 133.6-127.8 (C_{arom}), 91.8 (C₁, J_{C,H} = 167.0 Hz), 83.4, 79.2, 77.0, 73.1 (C₂/C₃/C₄/C₅), 74.9, 74.5, 71.9 (CH₂ Bn), 68.1 (C₆); β-anomer: δ 165.0 (C=O Bz), 138.1, 138.0, 137.9 (C_q Bn), 133.6-127.8 (C_{arom}), 93.5 (C₁, J_{C,H} = 160.0 Hz), 83.4, 77.0, 76.0, 72.9 (C₂/C₃/C₄/C₅), 74.9, 74.5, 71.9 (CH₂ Bn), 68.3 (C₆). MS (ESI): m/z 555 (M+H)⁺, 572 (M+NH₄)⁺, 577 (M+Na)⁺.

1,2-Di-*O*-benzoyl-3,4,6-tri-*O*-benzyl-α/β-D-glucopyranose (9). Yield: 0.58 g, 0.88 mmol, 88%, α:β = 1:11. α-anomer: 13 C NMR (CDCl₃): δ 164.9, 163.8 (C=O Bz), 138.1, 138.0, 137.9 (C_q Bn), 133.6-127.7 (C_{arom}), 91.2 (C₁, J_{C,H} = 168.2 Hz), 82.6, 79.1, 74.8, 72.7 (C₂/C₃/C₄/C₅), 75.3, 75.0, 73.5 (CH₂ Bn), 68.1 (C₆); β-anomer: δ 165.3, 165.0 (C=O Bz), 138.1, 138.0, 137.9 (C_q Bn), 133.6-127.7 (C_{arom}), 93.1 (C₁, J_{C,H} = 159.8 Hz), 82.6, 77.6, 76.1, 72.9 (C₂/C₃/C₄/C₅), 75.1, 75.0, 73.6 (CH₂ Bn), 68.3 (C₆). MS (ESI): m/z 659 (M+H)⁺, 676 (M+NH₄)⁺, 681 (M+Na)⁺.

Anal. Calcd for C₄₁H₃₈O₈ (658.1): C, 74.76; H, 5.81. Found: C, 74.58; H, 5.92.

1-*O*-Formyl-2-*O*-benzoyl-3,4,6-tri-*O*-benzyl-α/β-D-glucopyranose (11). Yield: 0.42 g, 0.72 mmol, 72%, α:β = 1:5. α-anomer: 13 C NMR (CDCl₃): δ 165.0 (C=O Bz), 161.3 (HC=O), 137.5, 137.4, 137.2 (C_q Bn), 134.0-127.1 (C_{arom}), 89.6 (C₁, J_{C,H} = 169.2 Hz), 78.0, 76.9, 73.2, 70.9 (C₂/C₃/C₄/C₅), 75.5, 74.2, 72.3 (CH₂ Bn), 67.9 (C₆); β-anomer: δ 161.7 (C=O Bz), 160.9 (HC=O), 137.5, 137.4, 137.2 (C_q Bn), 134.0-127.1 (C_{arom}), 91.4 (C₁, J_{C,H} = 158.0 Hz), 81.8, 76.8, 75.5, 71.9 (C₂/C₃/C₄/C₅), 74.9, 74.4, 71.7 (CH₂ Bn), 67.6 (C₆). MS (ESI): m/z 583 (M+H)⁺.

Methyl 2,3,4-Tri-*O*-benzyl-6-*O*-(3,4,6-tri-*O*-benzyl-α/β-D-glucopyranosyl)-α-D-glucuronopyranoside (12). Yield: 0.64 g, 0.70 mmol, 70%, α:β = 3:10. α-anomer: 13 C NMR (CDCl₃): δ 168.0 (C₆), 138.2, 138.0, 137.7, 137.5, 137.4, 137.4 (C_q Bn), 128.9-127.2 (C_{arom}), 98.1 (C₁, J_{C,H} = 169.8 Hz), 93.1 (C₁, J_{C,H} = 168.6 Hz), 84.2, 81.7, 80.7, 75.3, 72.5, 71.1, 70.0, 69.8 (C₂/C₃/C₄/C₅/C₂/C₃/C₄/C₅·), 75.2, 74.6, 74.2, 74.0, 72.8, 72.8 (CH₂ Bn), 67.6 (C₆·), 55.2 (OMe); β-anomer: δ 168.1 (C₆), 138.2, 138.0, 137.7, 137.5, 137.4, 137.4 (C_q Bn), 128.9-127.2 (C_{arom}), 98.1 (C₁, J_{C,H} = 169.8 Hz), 94.5 (C₁·, J_{C,H} = 158.9 Hz), 84.0, 80.7, 79.0, 78.8, 77.4, 76.6, 72.5, 69.8 (C₂/C₃/C₄/C₅/C₂·/C₃·/C₄·/C₅·), 75.2, 74.6, 74.2, 74.0, 72.8, 72.8 (CH₂ Bn), 68.0 (C₆·), 55.2 (OMe). MS (ESI): m/z 912 (M+H)⁺, 929 (M+NH₄)⁺, 934 (M+Na)⁺.

Anal. Calcd for C₅₅H₅₈O₁₂ (911.0): C, 72.51; H, 6.42. Found: C, 72.43; H, 6.45.

Dibenzyl-(3,4,6-tri-*O*-benzyl-β-D-glucopyranosyl) phosphate (13). Yield: 0.58 g, 0.82 mmol, 82%. 31 P NMR (CDCl₃): δ -2.01. 1 H NMR (CDCl₃): δ 7.58-7.06 (m, 25H, H_{arom}), 5.02 (t, 1H, H₁, J_{1,2} = J_{1,P} = 7.8 Hz), 4.84-4.41 (m, 10H, CH₂ Bn), 4.03 (dd, 1H, H₂, J_{2,3} = 8.6 Hz), 3.99 (dd, 1H, H₃, J_{3,4} = 7.7 Hz), 3.76 (m, 1H, H₅), 3.68-3.58 (m, 2H, H₄/H₆), 3.54 (dd, 1H, H₆, J_{5,6} = 4.8 Hz, J_{6,6} = 10.9 Hz), 2.60 (bs, 1H, OH). 13 C NMR (CDCl₃): δ 138.2, 138.1, 137.8 (C_q Bn sugar), 135.0 (C_q Bn phosph.), 128.6-127.2 (C_{arom}), 98.4 (C₁, J_{1,P} = 4.6 Hz), 83.9, 77.4, 75.0 (C₃/C₄/C₅), 75.2, 74.9, 73.9 (CH₂ Bn sugar), 74.8 (C₂, J_{2,P} = 7.2 Hz), 70.2, 69.9 (CH₂ Bn phosph., J_{C,P} = 4.0 Hz), 68.4 (C₆). MS (ESI): m/z 711 (M+H)⁺, 733 (M+Na)⁺.

Dibenzyl-(2-*O*-benzoyl-3,4,6-tri-*O*-benzyl-β-D-glucopyranosyl) phosphate (14). Yield: 0.59 g, 0.72 mmol, 72%. 31 P NMR (CDCl₃): δ -2.21. 1 H NMR (CDCl₃): δ 8.10-7.10 (m, 30H, H_{arom}), 5.40 (t, 1H, H₁, J_{1,2} = J_{1,P} = 7.6 Hz), 4.89 (dd, 1H, H₂, J_{2,3} = 8.6 Hz), 4.80-4.46 (m, 10H, CH₂ Bn), 4.06 (dd, 1H, H₃, J_{3,4} = 7.2 Hz), 3.90-3.62 (m, 4H, H₄/H₅/H₆/H₆·). 13 C NMR (CDCl₃): δ 165.1 (C=O Bz), 138.0, 137.9, 137.8 (C_q Bn sugar), 135.1 (C_q Bn phosph.), 133.2 (C_q Bz), 133.1-128.0 (C_{arom}), 96.2 (C₁, J_{1,P} = 4.8 Hz), 81.0, 76.8, 74.9 (C₃/C₄/C₅), 74.9, 74.8, 73.3 (CH₂ Bn sugar), 71.8 (C₂, J_{2,P} = 8.2 Hz), 70.1, 69.9 (CH₂ Bn phosph., J_{C,P} = 4.0 Hz), 68.4 (C₆). MS (ESI): m/z 815 (M+H)⁺, 832 (M+NH₄)⁺, 837 (M+Na)⁺.

Di-*n*-**butyl-**(3,4,6-tri-*O*-benzyl-β-D-glucopyranosyl) phosphate (15). Yield: 0.57 g, 0.88 mmol, 88%. ³¹P NMR (CDCl₃): δ -1.92. ¹H NMR (CDCl₃): δ 7.34-7.14 (m, 15H, H_{arom}), 5.03 (t, 1H, H₁, J_{1,2} = J_{1,P} = 7.8 Hz), 4.89 (AB, 2H, CH₂ Bn), 4.70 (AB, 2H, CH₂ Bn), 4.56 (AB, 2H, CH₂ Bn), 4.18 (dt, 4H, CH₂α Bu, J_{H,P} = 5.9 Hz), 4.10-3.99 (m, 2H, H₂/H₃), 3.70 (m, 1H, H₅), 3.62 (dd, 1H, H₄, J_{3,4} = 8.1 Hz, J_{4,5} = 7.7 Hz), 3.59 (dd, 1H, H₆, J_{5,6} = 2.9 Hz, J_{6,6} = 10.8 Hz), 3.52 (dd, 1H, H₆, J_{5,6} = 4.2 Hz), 2.95 (bs, 1H, OH), 1.60 (m, 4H, CH₂β Bu), 1.36 (m, 4H, CH₂γ Bu), 0.90 (t, 6H, CH₃ Bu). ¹³C NMR (CDCl₃): δ 138.4, 137.9, 137.8 (C_q Bn), 127.8-127.3 (C_{arom}), 99.0 (C₁, J_{1,P} = 4.4 Hz), 84.2, 76.8, 75.5 (C₃/C₄/C₅), 75.2, 74.8, 73.3 (CH₂ Bn), 74.6 (C₂, J_{2,P} = 7.3 Hz), 68.4 (C₆), 67.9 (CH₂α Bu, J_{C,P} = 4.9 Hz), 32.1 (CH₂β Bu, J_{C,P} = 5.8 Hz), 18.5 (CH₂γ Bu), 13.4 (CH₃ Bu). MS (ESI): *m/z* 643 (M+H)⁺, 665 (M+Na)⁺.

Di-*n*-butyl-(2-*O*-benzoyl-3,4,6-tri-*O*-benzyl-β-D-glucopyranosyl) phosphate (16). Yield: 0.60 g, 0.81 mmol, 81%. ³¹P NMR (CDCl₃): δ -2.10. ¹H NMR (CDCl₃): δ 8.05-7.11 (m, 20H, H_{arom}), 5.35 (dd, 1H, H₁, J_{1,2} = 7.9 Hz, J_{1,P} = 6.0 Hz), 4.80 (dd, 1H, H₂, J_{2,3} = 8.4 Hz), 4.76 (AB, 2H, CH₂ Bn), 4.68 (AB, 2H, CH₂ Bn), 4.52 (AB, 2H, CH₂ Bn), 4.20 (dt, 4H, CH₂α Bu, J_{H,P} = 6.2 Hz), 4.00 (dd, 1H, H₃, J_{3,4} = 7.9 Hz), 3.85-3.60 (m, 4H, H₄/H₅/H₆/H₆·), 1.50 (m, 4H, CH₂β Bu), 1.31 (m, 4H, CH₂γ Bu), 0.90 (t, 6H, CH₃ Bu). ¹³C NMR (CDCl₃): δ 164.9 (C=O Bz), 137.7, 137.6, 137.4 (C_q Bn), 133.6-127.5 (C_{arom}), 132.9 (C_q Bz), 96.5 (C₁, J_{1,P} = 4.0 Hz), 81.9, 77.3, 75.5 (C₃/C₄/C₅), 74.9, 74.8, 73.3 (CH₂ Bn), 73.4 (C₂, J_{2,P} = 7.8 Hz), 68.1 (C₆), 67.6 (CH₂α Bu, J_{C,P} = 4.2 Hz), 31.6 (CH₂β Bu, J_{C,P} = 7.4 Hz), 18.4 (CH₂γ Bu), 13.3 (CH₃ Bu). MS (ESI): *m/z* 747 (M+H)⁺, 769 (M+Na)⁺.

Methyl 2,3,4-Tri-*O*-benzyl-6-[benzyl-(3,4,6-tri-*O*-benzyl-α/β-D-glucopyranosyl) phosphate]-α-D-glucopyranoside (17). Yield: 0.79 g, 0.74 mmol, 74%, α:β = 1:4, $R_p:S_p=1:1.\ ^{13}C$ NMR (CDCl₃): α-anomer: δ 138.4-135.9 (C_q Bn), 128.1-127.7 (C_{arom}), 97.8 (C_1 ', $J_{C,H}=167.3$ Hz, $J_{C,P}=4.4$ Hz), 97.7 (C_1 , $J_{C,H}=169.2$ Hz), 84.5-70.1 ($C_2/C_3/C_4/C_5/C_2/C_3/C_4/C_5$), 75.4-73.1 (CH₂ Bn sugar), 69.4 (C_6), 68.3 (CH₂ Bn phosph., $J_{C,P}=4.4$ Hz), 65.3 (C_6 , $J_{C,P}=4.9$ Hz), 55.0 (OMe); β-anomer: δ 138.4-137.8 (C_q Bn), 128.1-127.7 (C_{arom}), 98.9 (C_1 ', $J_{C,H}=160.0$ Hz, $J_{C,P}=4.5$ Hz), 97.7 (C_1 , $J_{C,H}=169.2$ Hz), 83.9-69.3 ($C_2/C_3/C_4/C_5/C_2/C_3/C_4/C_5$), 75.4-73.1 (CH₂ Bn sugar), 69.4 (C_6), 68.5 (CH₂ Bn phosph., $J_{C,P}=4.4$ Hz), 65.7 (C_6 , $J_{C,P}=5.0$ Hz), 55.0 (OMe). MS (ESI): m/z 1068 (M+H)⁺, 1085 (M+NH₄)⁺, 1090 (M+Na)⁺.

1,5-Anhydro-2-deoxy-3,4-di-O-benzyl-6-O-tert-butyldiphenylsilyl-D-arabino-hex-1-enitol (18). A solution of 6-O-tert-butyldiphenylsilyl-D-glucal (3.84 g, 10.0 mmol) in THF (30 mL) was heated to 30 °C. To the latter solution, NaH (60% dispersion in mineral oil, 2.4 g, 60 mmol) was added and the mixture was stirred for 10 min. Subsequently methyltriphenylphosphonium iodide (8.1 g, 20 mmol) and benzyl bromide

(7.2 mL, 60 mmol) were added and the reaction mixture was stirred at 30 °C for 3 h. Methanol (3 mL) was added and the heterogeneous mixture was concentrated *in vacuo*. The residue was dissolved in diethyl ether (200 mL), washed with sat. aq. NaCl (3 x 50 mL), dried (MgSO₄) and concentrated under reduced pressure. Purification of the residue was effected by silica gel chromatography (0-10% EtOAc/light petroleum) to give fully protected glucal **18** (4.62 g, 8.2 mmol, 82%) as a white solid. ¹H NMR (CDCl₃): δ 7.74-7.24 (m, 20H, H_{arom}), 6.40 (dd, 1H, H₁, J_{1,2} = 5.9 Hz, J_{1,3} = 1.3 Hz), 4.84 (dd, 1H, H₂, J_{2,3} = 2.6 Hz), 4.82 (AB, 2H, CH₂ Bn), 4.61 (AB, 2H, CH₂ Bn), 4.21 (ddd, 1H, H₃, J_{3,4} = 7.2 Hz), 4.00 (t, 1H, H₄, J_{4,5} = 7.3 Hz), 3.97-3.91 (m, 3H, H₅/H₆/H₆), 1.06 (s, 9H, CH₃ t-Bu). ¹³C NMR (CDCl₃): δ 144.9 (C₁), 138.6, 138.5 (C_q Bn), 136.0-127.8 (C_{arom}), 133.7, 133.3 (C_q TBDPS), 99.8 (C₂), 78.1, 76.1, 74.3 (C₃/C₄/C₅), 74.0, 70.7 (CH₂ Bn), 62.4 (C₆), 27.0 (CH₃ t-Bu), 19.5 (C_q t-Bu).

Anal. Calcd for $C_{36}H_{40}O_4Si$ (564.5): C, 76.56; H, 7.14; Si, 4.97. Found: C, 76.50; H, 7.21; Si, 4.98.

1,2-Anhydro-3,4-di-*O*-benzyl-6-*O*-tert-butyldiphenylsilyl-α-D-glucopyranose (19). To a stirred and cooled (0 °C) solution of glucal 18 (2.82 g, 5.0 mmol) in CH₂Cl₂ (10 mL) was added a freshly prepared solution of 3,3-dimethyldioxirane (DMD, 67 mL, 0.09 M, 6.0 mmol) in acetone. Immediately after the last addition, the reaction mixture was concentrated under reduced pressure to afford epoxide 19 as a white solid in quantitative yield (2.90 g, 5.0 mmol). ¹H NMR (CDCl₃): δ 7.81-7.18 (m, 20H, H_{arom}), 4.96 (d, 1H, H₁, J_{1,2} = 3.4 Hz), 4.76 (AB, 2H, CH₂ Bn), 4.58 (AB, 2H, CH₂ Bn), 3.99 (dd, 1H, H₃, J_{2,3} = 1.4 Hz, J_{3,4} = 7.9 Hz), 3.80 (m, 1H, H₅), 3.60 (dd, 1H, H₆, J_{5,6} = 2.9 Hz, J_{6,6} = 10.8 Hz), 3.58 (dd, 1H, H₄, J_{4,5} = 7.4 Hz), 3.49 (dd, 1H, H₆, J_{5,6} = 3.6 Hz), 3.12 (dd, 1H, H₂), 1.04 (s, 9H, CH₃ t-Bu). ¹³C NMR (CDCl₃): δ 138.8, 137.9 (C_q Bn), 136.1-127.7 (C_{arom}), 133.8, 133.2 (C_q TBDPS), 79.4, 74.4, 70.7 (C₃/C₄/C₅), 79.0 (C₁), 74.8, 70.7 (CH₂ Bn), 62.3 (C₆), 52.7 (C₂), 27.1 (CH₃ t-Bu), 19.5 (C_q t-Bu).

1-*O*-Benzoyl-3,4-di-*O*-benzyl-6-*O*-tert-butyldiphenylsilyl-β-D-glucopyranose (20). Anomeric benzoate 20 (3.23 g, 4.6 mmol, 92%) was prepared from 1,2-anhydroglucose derivative 19 (2.90 g, 5.0 mmol) as described in the general procedure for the conversion of epoxide 6. ¹H NMR (CDCl₃): δ 8.12-7.12 (m, 25H, H_{arom}), 5.81 (d, 1H, H₁, J_{1,2} = 7.5 Hz), 4.92 (AB, 2H, CH₂ Bn), 4.80 (AB, 2H, CH₂ Bn), 4.04 (t, 1H, H₂, J_{2,3} = 7.6 Hz), 4.00-3.78 (m, 3H, H₃/H₄/H₅), 3.70 (dd, 1H, H₆, J_{5,6} = 2.0 Hz, J_{6,6} = 10.9 Hz), 3.52 (dd, 1H, H₆, J_{5,6} = 4.1 Hz), 2.60 (bs, 1H, OH), 1.04 (s, 9H, CH₃ t-Bu). ¹³C NMR (CDCl₃): δ 164.8 (C=O Bz), 138.2, 138.0 (C_q Bn), 135.5-127.2 (C_{arom}), 133.2, 132.6 (C_q TBDPS), 129.0 (C_q Bz), 94.5 (C₁), 84.2, 76.6, 76.0, 73.1 (C₂/C₃/C₄/C₅), 75.0, 74.7 (CH₂ Bn), 62.1 (C₆), 26.5 (CH₃ t-Bu), 19.0 (C_q t-Bu). MS (ESI): m/z 703 (M+H)⁺, 725 (M+Na)⁺.

Anal. Calcd for $C_{43}H_{46}O_7Si$ (702.3): C, 73.48; H, 6.60; Si, 4.00. Found: C, 73.40; H, 6.52; Si, 4.12.

1-O-Benzoyl-2-O-(2,3,4,6-tetra-O-benzyl- α -D-glucopyranosyl)-3,4-di-O-benzyl-6-O-tert-butyldiphenylsilyl-β-D-glucopyranose (22). A mixture of anomeric benzoate 20 (1.40 g, 2.0 mmol), thioethyl donor 21 (1.40 g, 2.4 mmol) and powdered molecular sieves (4 Å, 0.3 g) in 1,2-dichloroethane/diethyl ether (10 mL, 1:1, v/v) was stirred under a continuous stream of dry nitrogen. After 30 min, IDCT (1.55 g, 3.0 mmol) was added in one portion and the resulting mixture was stirred for 1 h, subsequently diluted with diethyl ether (100 mL) and washed with aq. Na₂S₂O₃ (1.0 M, 2 x 25 mL) and aq. NaHCO3 (1.0 M, 2 x 25 mL). The organic layer was dried (MgSO4) and concentrated in vacuo. The resulting yellow oil was purified by silica gel chromatography (10-30% EtOAc/light petroleum) and LH-20 gel filtration (eluent: CH₂Cl₂/MeOH, 2:1, v/v) to give dimer 22 as a white solid (1.76 g, 1.44 mmol, 72%). ¹H NMR (CDCl₃): δ 8.10-7.00 (m, 45H, H_{arom}), 6.06 (d, 1H, H_1 , $J_{1,2} = 8.1$ Hz), 5.54 (d, 1H, $H_{1'}$, $J_{1',2'} = 3.6$ Hz), 4.91 (AB, 2H, CH₂ Bn), 4.82 (AB, 2H, CH₂ Bn), 4.80 (AB, 2H, CH₂ Bn), 4.62 (AB, 2H, CH₂ Bn), 4.40 (AB, 2H, CH₂ Bn), 4.31 (AB, 2H, CH₂ Bn), 4.11 (dd, 1H, H₂, $J_{2,3} =$ 9.0 Hz), 4.05-3.93 (m, 3H, $H_4/H_5/H_{3'}$), 3.87 (t, 1H, H_3 , $J_{3,4} = 9.2$ Hz), 3.66 (dd, 1H, $H_{4'}$, $J_{3',4'} = 9.8 \text{ Hz}, J_{4',5'} = 9.4 \text{ Hz}), 3.57 \text{ (dd, 1H, H}_{6A}, J_{5,6A} = 2.0 \text{ Hz}, J_{6A,6B} = 9.5 \text{ Hz}), 3.51$ (dd, 1H, $H_{2'}$, $J_{2',3'}$ = 9.7 Hz), 3.45 (dd, 1H, H_{6B} , $J_{5,6B}$ = 4.2 Hz), 3.37 (dd, 1H, $H_{6'A}$, $J_{5',6'A}$ = 1.8 Hz, $J_{6'A.6'B}$ = 10.9 Hz), 3.25 (dd, 1H, $H_{6'B}$, $J_{5'.6'B}$ = 2.3 Hz), 1.04 (s, 9H, CH₃ t-Bu). 13 C NMR (CDCl₃): δ 164.4 (C=O Bz), 138.6, 138.6, 138.0, 137.9, 137.7, 137.6 (C_q Bn), 135.8-127.3 (C_{arom}), 133.4, 132.7 (C_q TBDPS), 129.2 (C_q Bz), 95.8, 95.2 (C₁/C₁), 82.9, $81.8,\ 78.8,\ 78.1,\ 77.4,\ 76.0,\ 74.0,\ 69.9\ (C_2/C_3/C_4/C_5/C_2/C_3/C_4/C_5),\ 75.5,\ 75.0,\ 74.8,$ 73.3, 73.2, 72.3 (CH₂ Bn), 67.7 (C₆), 61.9 (C₆), 26.7 (CH₃ t-Bu), 19.3 (C_q t-Bu). MS (ESI): m/z 1226 (M+H)⁺.

Anal. Calcd for $C_{77}H_{80}O_{12}Si$ (1225.1): C, 75.46; H, 6.58; Si, 2.29. Found: C 75.38; H, 6.58; Si, 2.27.

1,2-Di-O-tert-butyldiphenylsilyl-sn-glycerol (23). To a stirred solution of commercially available 1,2-O-isopropylidene-sn-glycerol (2.49 mL, 20 mmol) in DMF (100 mL) were added NaH (60% dispersion in mineral oil, 0.96 g, 24 mmol) and BnBr (2.85 mL, 24 mmol). After 1 h, the reaction mixture was quenched by addition of MeOH (3 mL) and concentrated in vacuo. The residue was taken up in diethyl ether (200 mL), washed with aq. NaCl (1.0 M, 2 x 50 mL), dried (MgSO₄) and concentrated under reduced pressure. The resulting pale yellow oil was dissolved in AcOH (40 mL) and water (10 mL), heated under reflux for 1 h and subsequently concentrated in vacuo. Residual AcOH was removed by coevaporation with dioxane (3 x 50 mL), after which pyridine (100 mL) and TBDPSCl (13.0 mL, 50 mmol) were sequentially added. The

reaction mixture was stirred for 12 h and then quenched by addition of MeOH (3 mL). The resulting solution was concentrated under reduced pressure, dissolved in diethyl ether (200 mL) and washed with aq. NaHCO₃ (1.0 M, 3 x 50 mL), dried (MgSO₄) and concentrated *in vacuo*. Traces of pyridine in the residue were removed by coevaporation with toluene (3 x 50 mL). The thus obtained yellow oil was dissolved in ethyl acetate (50 mL) and MeOH (10 mL), upon which palladium on charcoal (10% Pd, 500 mg) was added. The latter heterogeneous mixture was hydrogenated at elevated pressure (3 atm.) in a Parr apparatus for 12 h. The metal catalyst was removed by filtration and the filtrate was concentrated under reduced pressure. Purification of the residue was effected by silica gel chromatography (0-15% EtOAc/light petroleum) to give glycerol **23** as a colorless oil (7.72 g, 13.6 mmol, 68%). [α]_D = -24.8 ° (c 1.2, CHCl₃). ¹H NMR (CDCl₃): δ 7.70-7.24 (m, 20H, H_{arom}), 3.86 (dd, 1H, H₁, J_{1,1'} = 11.4 Hz, J_{1,2} = 4.9 Hz), 3.82 (dd, 1H, H_{1'}, J_{1',2} = 3.0 Hz), 3.72-3.65 (m, 2H, H₂/H₃), 3.58 (m, 1H, H_{3'}), 1.02, 0.98 (2 x s, 2 x 9H, CH₃ t-Bu). ¹³C NMR (CDCl₃): δ 135.5-127.5 (C_{arom}), 133.3, 132.9 (C_q TBDPS), 73.4 (C₂), 64.7 (C₁), 63.9 (C₃), 26.7, 26.6 (CH₃ t-Bu), 19.9, 19.0 (C_q t-Bu).

1,2-Di-O-tert-butyldiphenylsilyl-3-O-(3,4-di-O-benzyl-6-O-tert-butyldiphenylsilyl-2-O-(2,3,4,6-tetra-O-benzyl- α -D-glucopyranosyl)- α/β -D-glucopyranosyl)-sn-glycerol (25). A mixture of dimer benzoate 22 (0.61 g, 0.5 mmol), glycerol 23 (0.34 g, 0.6 mmol) and powdered molecular sieves (4 Å, 0.1 g) in 1,2-dichloroethane (3 mL) was stirred under a continuous stream of dry nitrogen for 15 min. To the latter mixture, TMSOTf was added in three portions at 5 min-intervals (3 x 48 µL, 3 x 0.25 mmol). Stirring was continued for 3 h and the reaction mixture was neutralized by addition of triethylamine (1 mL), diluted with diethyl ether (50 mL) and washed with aq. NaHCO₃ (3 x 25 mL). The organic layer was dried (MgSO₄) and concentrated under reduced pressure. Purification of the residue was effected with silica gel chromatography (0-15% EtOAc/light petroleum) to furnish 25 as a mixture of anomers (colorless oil, 0.68 g, 0.41 mmol, 81%, α : β = 8:1). ¹³C NMR (CDCl₃): α -anomer: δ 139.8, 139.6, 139.0, 138.7, 138.4, 138.2 (C_q Bn), 137.9-129.0 (C_{arom}), 136.2, 135.8, 135.6, 135.4, 135.3, 135.1 (C_q TBDPS), 96.5 ($C_{1'}$, $J_{C,H} = 168.4$ Hz), 94.5 ($C_{1''}$, $J_{C,H} = 170.0$ Hz), 82.1, 81.1, 79.2, 77.8, 77.4, 74.4, 72.4, 71.3, 70.0 $(C_2/C_2/C_3/C_4/C_5/C_2^n/C_3^n/C_4^n/C_5^n)$, 75.5, 75.4, 74.7, 73.3, 73.1, 71.5 (CH₂ Bn), 67.8 ($C_{6''}$), 65.4 (C_3), 62.7, 62.1 ($C_1/C_{6'}$), 25.2, 25.1, 24.9 (CH₃ t-Bu), 17.3, 17.2, 17.1 (C_q *t*-Bu); β-anomer: δ 139.7, 139.6, 139.1, 138.7, 138.4, 138.1 (C_q Bn), 137.9-129.0 (C_{arom}), 136.2, 135.8, 135.6, 135.4, 135.3, 135.1 (C_q TBDPS), 99.3 $(C_{1'}, J_{C,H} = 159.9 \text{ Hz}), 94.9 (C_{1''}, J_{C,H} = 170.6 \text{ Hz}), 81.9, 80.0, 79.0, 77.8, 77.3, 74.5,$ (CH₂ Bn), 67.9 (C_{6"}), 64.7 (C₃), 63.2, 62.1 (C₁/C₆), 25.2, 25.1, 24.9 (CH₃ t-Bu), 17.3, 17.2, 17.1 (C_q t-Bu). MS (ESI): m/z 1673 (M+H)⁺, 1690 (M+NH₄)⁺, 1695 (M+Na)⁺.

Anal. Calcd for $C_{105}H_{118}O_{13}Si_3$ (1672.2): C, 75.41; H, 7.11; Si, 5.04. Found: C, 75.40; H, 7.16; Si, 5.00.

1,2-Di-O-allyl-3-O-(3,4-di-O-benzyl-6-O-tert-butyldiphenylsilyl-2-O-(2,3,4,6tetra-O-benzyl- α -D-glucopyranosyl)- α/β -D-glucopyranosyl)-sn-glycerol mixture of dimer benzoate 22 (0.61 g, 0.5 mmol), glycerol 24 (0.10 g, 0.6 mmol) and powdered molecular sieves (4 Å, 0.1 g) in 1,2-dichloroethane (3 mL) was stirred under a continuous stream of dry nitrogen for 15 min. To the latter mixture, TMSOTf was added in three portions at 5 min-intervals (3 x 48 µL, 3 x 0.25 mmol). Stirring was continued for 3 h and the reaction mixture was neutralized by addition of triethylamine (1 mL), diluted with EtOAc (50 mL) and washed with aq. NaHCO₃ (1.0 M, 3 x 25 mL). The organic layer was dried (MgSO₄) and concentrated under reduced pressure. Purification of the residue was effected with silica gel chromatography (10-25% EtOAc/light petroleum) to furnish anomeric mixture 26 as a white solid (0.47 g, 0.37 mmol, 74%, α : β = 6:1). ¹³C NMR (CDCl₃): α-anomer: δ 138.6, 138.5, 138.3, 138.1, 137.7, 137.6 (C_q Bn), 134.7 (CH All), 133.8-127.5 (C_{arom}), 133.4, 133.0 (C_q TBDPS), 116.7 (CH₂ All), 95.6 $(C_{1'}, J_{C,H} = 171.2 \text{ Hz}), 94.5 (C_{1''}, J_{C,H} = 169.4 \text{ Hz}), 81.9, 80.8, 79.0, 78.4, 77.8, 77.4,$ 76.5, 75.7, 71.5 $(C_2/C_2/C_3/C_4/C_5/C_2/C_3/C_4/C_5)$, 76.1, 75.5, 74.8, 73.2, 72.1, 72.0, 71.2, 70.8 (6 x CH₂ Bn, 2 x CH₂ All), 69.9, 67.9, 67.1 (C₁/C₃/C_{6"}), 62.5 (C_{6'}), 26.7 (CH₃ t-Bu), 19.2 (C_q t-Bu); β -anomer: δ 138.6, 138.5, 138.3, 138.1, 137.7, 137.6 (C_q Bn), 134.7 (CH All), 133.8-127.5 (C_{arom}), 133.4, 133.0 (C_q TBDPS), 116.8 (CH₂ All), 103.5 $(C_{1'}, J_{C.H} = 161.8 \text{ Hz}), 94.7 (C_{1''}, J_{C.H} = 169.6 \text{ Hz}), 82.7, 80.8, 79.0, 78.9, 78.4, 77.8,$ 70.4, 70.3 (6 x CH₂ Bn, 2 x CH₂ All), 69.9, 67.8, 67.1 (C₁/C₃/C_{6"}), 62.2 (C_{6'}), 26.7 (CH₃ t-Bu), 19.2 (C_q t-Bu). MS (ESI): m/z 1276 (M+H)⁺, 1298 (M+Na)⁺.

1,2-Di-*O*-allyl-3-*O*-(3,4-di-*O*-benzyl-2-*O*-(2,3,4,6-tetra-*O*-benzyl-α-D-glucopyranosyl)- α -D-glucopyranosyl)-sn-glycerol (27). To a stirred solution of trimer 26 (0.47 g, 0.37 mmol) in THF (3 mL) was added tetra-n-butylammonium fluoride (1.0 M solution in THF, 0.5 mL). The reaction mixture was stirred for 2 h, subsequently diluted with EtOAc (50 mL) and washed with sat. aq. NaCl (3 x 25 mL). The organic layer was dried (MgSO₄) and concentrated *in vacuo*. Purification of the residue was effected with silica gel chromatography (10-50% EtOAc/light petroleum) to furnish primary alcohol 27 as a white solid (0.32 g, 0.31 mmol, 83% based on anomeric mixture 26). ¹H NMR (CDCl₃): δ 7.38-7.03 (m, 30H, H_{arom}), 5.92 (m, 2H, CH All), 5.27, 5.21, 5.16, 5.09 (4 x dd, 4 x 1H, CH₂ All), 5.06 (d, 1H, H_{1"}, J_{1",2"} = 3.5 Hz), 5.03 (d, 1H, H_{1'}, J_{1',2'} = 3.4 Hz), 4.91 (AB, 2H, CH₂ Bn), 4.87 (AB, 2H, CH₂ Bn), 4.76 (AB, 2H, CH₂ Bn), 4.60 (AB, 2H, CH₂ Bn), 4.54 (AB, 2H, CH₂ Bn), 4.46 (AB, 2H, CH₂ Bn), 4.07 (m, 2H, CH₂O All), 4.05 (t, 1H, H_{3"}, J_{2",3"} = J_{3",4"} = 8.4 Hz), 4.02 (m, 1H, H₂), 3.99 (dd, 1H, H_{3'}, J_{2',3'} = 9.0 Hz, J_{3',4'} = 8.2

Hz), 3.97-3.90 (m, 6H, $H_{1A}/H_{1B}/H_{3A}/H_{3B}/CH_2O$ All), 3.80 (dt, 1H, $H_{5'}$, $J_{4',5'} = 9.7$ Hz, $J_{5',6'A} = J_{5',6'B} = 4.8$ Hz), 3.76 (dd, 1H, $H_{2'}$), 3.74 (m, 1H, $H_{5''}$), 3.66 (dd, 1H, $H_{4''}$, $J_{4'',5''} = 9.0$ Hz), 3.63 (dd, 1H, $H_{6'A}$, $J_{6'A,6'B} = 10.8$ Hz), 3.57 (dd, 1H, $H_{2''}$), 3.53 (m, 2H, $H_{6''A}/H_{6''B}$), 3.50 (dd, 1H, $H_{4'}$), 3.46 (dd, 1H, $H_{6'B}$), 1.70 (bs, 1H, OH). ¹³C NMR (CDCl₃): δ 138.6, 138.1, 138.0, 137.9, 137.8, 137.7 (C_q Bn), 135.0, 134.6 (CH All), 128.8-127.8 (C_{arom}), 116.9, 116.8 (CH₂ All), 95.8, 94.5 ($C_{1'}/C_{1''}$), 81.9, 80.5, 79.0, 77.5, 77.4, 76.5, 75.4, 71.0, 70.3 ($C_2/C_2/C_3/C_4/C_5/C_2/C_3/C_4/C_5/C_2/C_3/C_4/C_5/C_2/C_3/C_4/C_5/C_2/C_3/C_4/C_5/C_2/C_3/C_4/C_5/C_2/C_3/C_4/C_5/C_2/C_3/C_4/C_5/C_2/C_3/C_4/C_5/C_5/C_3/C_4/C_5/C_5/C_3/C_4/C_5/C_5/C_3/C_4/C_5/C_5/C_3/C_4/C_5/C_5/C_3/C_4/C_5/C_5/C_3/$

3-O-(3,4-Di-O-benzyl-2-O-(2,3,4,6-tetra-O-benzyl- α -D-glucopyranosyl)- α -Dglucopyranosyl)-sn-glycerol (28). Tetra-n-butylammonium fluoride (1.0 M solution in THF, 2.0 mL) was added to a stirred solution of trimer 25 (0.68 g, 0.41 mmol) in THF (5 mL). The reaction mixture was stirred for 4 h, subsequently diluted with EtOAc (50 mL) and washed with sat. aq. NaCl (3 x 25 mL). The organic layer was dried (MgSO₄) and concentrated in vacuo. Purification of the residue was accomplished by silica gel chromatography (10-80% EtOAc/light petroleum) to afford triol 28 as a white solid (0.31 g, 0.32 mmol, 78% based on anomeric mixture 25). ¹H NMR (CDCl₃): δ 7.40-7.05 (m, 30H, H_{arom}), 4.95 (d, 1H, $H_{1'}$, $J_{1',2'} = 3.4$ Hz), 4.90 (d, 1H, $H_{1''}$, $J_{1'',2''} = 3.9$ Hz), 4.85 (AB, 2H, CH₂ Bn), 4.70 (AB, 2H, CH₂ Bn), 4.62 (AB, 2H, CH₂ Bn), 4.58 (AB, 2H, CH₂ Bn), 4.50 (AB, 2H, CH₂ Bn), 4.48 (AB, 2H, CH₂ Bn), 4.06 (dd, 1H, H_{3"}, J_{2",3"} = 7.9 Hz, J_{3",4"} = 8.9 Hz), 4.00 (t, 1H, $H_{3'}$, $J_{2',3'}$ = $J_{3',4'}$ = 9.1 Hz), 3.97 (m, 2H, H_{3A}/H_{3B}), 3.92 (m, 1H, $H_{5"}$), 3.82 (dd, 1H, $H_{6"A}$, $J_{5",6"A}$ = 4.0 Hz, $J_{6"A,6"B}$ = 10.7 Hz), 3.75 (dd, 1H, $H_{6"B}$, $J_{5",6"B}$ = 5.3 Hz), 3.71 (dd, 1H, H₂), 3.67-3.63 (m, 3H, H₅/H_{1A}/H_{1B}), 3.61 (dd, 1H, H_{4"}, J_{4".5"} = 9.2 Hz), 3.57 (dd, 1H, $H_{2"}$), 3.50 (dd, 1H, $H_{4'}$, $J_{4',5'}$ = 8.4 Hz), 3.40 (dd, 1H, $H_{6'A}$, $J_{5',6'A}$ = 3.9 Hz, $J_{6'A,6'B} = 10.8$ Hz), 3.33 (m, 1H, H₂), 3.30 (dd, 1H, H_{6'B}, $J_{5',6'B} = 5.6$ Hz). ¹³C NMR (CDCl₃): 8 138.4, 138.2, 138.0, 138.0, 137.9, 137.8 (C_a Bn), 128.4-127.6 (C_{arom}), 96.0, 94.9 $(C_1/C_{1'})$, 81.9, 80.5, 79.3, 78.1, 77.8, 77.5, 77.4, 71.3, 70.4 $(C_2/C_2/C_3/C_4/C_5/C_2/C_3/C_3/C_4/C_5)$, 75.9, 75.5, 75.1, 74.8, 73.3, 72.7 (CH₂ Bn), 71.6, 68.1, 68.0 ($C_1/C_3/C_{6''}$), 61.8 ($C_{6'}$).

1,2-Di-O-palmitoyl-3-O-(3,4-di-O-benzyl-6-O-palmitoyl-2-O-(2,3,4,6-tetra-O-benzyl- α -D-glucopyranosyl)- α -D-glucopyranosyl)-sn-glycerol (29). Palmitoyl chloride (0.11 mL, 0.36 mmol) was added dropwise to a stirred solution of triol 28 (95 mg, 0.10 mmol) in pyridine/CH₂Cl₂ (3 mL, 1:1, v/v). After 3 h, the reaction mixture was quenched by addition of H₂O (0.5 mL) and the solvents were removed by evaporation in vacuo. The residue was taken up in diethyl ether (25 mL) and washed thoroughly with aq. NaHCO₃ (1.0 M, 4 x 10 mL). The organic phase was dried (MgSO₄) and concentrated under reduced pressure. Traces of pyridine in the residue were removed by coevaporation with toluene (3 x 10 mL). Further purification was accomplished by LH-20 gel filtration

(eluent: CH₂Cl₂/MeOH, 2:1, v/v) to give fully protected glycolipid 29 as a greasy solid (154 mg, 0.092 mmol, 92%). ¹H NMR (CDCl₃): 8 7.36-7.05 (m, 30H, H_{arom}), 5.26 (m, 1H, H₂), 5.16 (d, 1H, H_{1"}, $J_{1",2"} = 3.1$ Hz), 5.14 (d, 1H, H_{1'}, $J_{1',2'} = 3.3$ Hz), 4.92 (AB, 2H, CH₂ Bn), 4.87 (AB, 2H, CH₂ Bn), 4.82 (AB, 2H, CH₂ Bn), 4.70 (AB, 2H, CH₂ Bn), 4.62 (AB, 2H, CH₂ Bn), 4.49 (AB, 2H, CH₂ Bn), 4.34 (dd, 1H, H_{1A} , $J_{1A,1B}$ = 12.1 Hz, $J_{1A,2}$ = 3.2 Hz), 4.28 (d, 2H, $H_{6'A}/H_{6'B}$, $J_{5'.6'} = 4.4$ Hz), 4.20 (dd, 1H, H_{1B} , $J_{1B,2} = 6.3$ Hz), 4.04 (t, 1H, $H_{3"}$, $J_{2",3"} = J_{3",4"} = 9.3$ Hz), 3.98 (t, 1H, $H_{3'}$, $J_{2',3'} = J_{3',4'} = 8.9$ Hz), 3.95 (m, 1H, $H_{5"}$), 3.85 (dt, 1H, $H_{5'}$, $J_{4',5'} = 10.0$ Hz), 3.75 (dd, 1H, $H_{2'}$), 3.72 (dd, 1H, H_{3A} , $J_{2.3A} = 3.9$ Hz, $J_{3A,3B} = 10.9 \text{ Hz}$), 3.69 (t, 1H, $H_{4"}$, $J_{4",5"} = 9.4 \text{ Hz}$), 3.60 (dd, 1H, H_{3B} , $J_{2,3B} = 3.1 \text{ Hz}$), 3.58 (dd, 1H, $H_{2"}$), 3.48 (dd, 1H, $H_{4'}$), 3.46 (dd, 1H, $H_{6"A}$, $J_{5".6"A} = 2.9$ Hz, $J_{6"A.6"B} = 9.2$ Hz), 3.37 (dd, 1H, $H_{6"B}$, $J_{5".6"B}$ = 3.5 Hz), 2.37-2.22 (m, 6H, 3 x $CH_2\alpha$ palm.), 1.75-0.87 (m, 87H, CH_2/CH_3 palm.). ¹³C NMR (CDCl₃): δ 173.2, 173.1, 172.7 (C=O palm.), 138.6, 138.4, 138.1, 137.9, 137.7, 137.6 (C_q Bn), 128.3-127.3 (C_{arom}), 96.2, 95.3 (C₁//C₁), 81.9, 80.4, 79.1, 77.7, 77.5, 75.9, 70.4, 69.7, 69.1 $(C_2/C_2/C_3/C_4/C_5/C_2/C_3/C_4/C_5/C_2/C_3/C_4/C_5)$, 75.5, 75.0, 74.8, 73.2, 73.1, 72.9 (CH₂ Bn), 67.9 (C_{6"}), 66.3 (C₃), 62.5, 62.4 (C₁/C_{6'}), 34.0-22.6 (CH₂ palm.), 14.0 (CH₃ palm.).

1,2-Di-O-myristoyl-3-O-(3,4-di-O-benzyl-6-O-myristoyl-2-O-(2,3,4,6-tetra-Obenzyl- α -D-glucopyranosyl)- α -D-glucopyranosyl)-sn-glycerol (30). To a stirred solution of triol 28 (95 mg, 0.10 mmol) in pyridine/CH₂Cl₂ (3 mL, 1:1, v/v) was added dropwise myristoyl chloride (0.098 mL, 0.36 mmol). After 3 h, the reaction mixture was quenched by addition of H₂O (0.5 mL) and the solvents were removed by evaporation in vacuo. The residue was taken up in diethyl ether (25 mL) and washed thoroughly with aq. NaHCO₃ (1.0 M, 4 x 10 mL). The organic phase was dried (MgSO₄) and concentrated under reduced pressure. Traces of pyridine in the residue were removed by coevaporation with toluene (3 x 10 mL). Further purification was accomplished by LH-20 gel filtration (eluent: CH2Cl2/MeOH, 2:1, v/v) to give fully protected glycolipid 30 as a greasy solid (143 mg, 0.090 mmol, 90%). ¹H NMR (CDCl₃): δ 7.45-7.04 (m, 30H, H_{arom}), 5.24 (m, 1H, H₂), 5.16 (d, 1H, H_{1"}, $J_{1",2"} = 3.1$ Hz), 5.12 (d, 1H, H_{1'}, $J_{1',2'} = 3.3$ Hz), 4.90 (AB, 2H, CH₂ Bn), 4.87 (AB, 2H, CH₂ Bn), 4.81 (AB, 2H, CH₂ Bn), 4.70 (AB, 2H, CH₂ Bn), 4.62 (AB, 2H, CH₂ Bn), 4.43 (AB, 2H, CH₂ Bn), 4.30 (dd, 1H, H_{1A} , $J_{1A,1B} = 12.1$ Hz, $J_{1A,2} = 12.1$ 3.2 Hz), 4.29 (d, 2H, $H_{6'A}/H_{6'B}$, $J_{5'.6'} = 4.4$ Hz), 4.18 (dd, 1H, H_{1B} , $J_{1B,2} = 6.3$ Hz), 4.02 (t, 1H, $H_{3"}$, $J_{2",3"} = J_{3",4"} = 9.3$ Hz), 3.99 (t, 1H, $H_{3'}$, $J_{2',3'} = J_{3',4'} = 8.9$ Hz), 3.96 (m, 1H, $H_{5"}$), 3.85 (dt, 1H, $H_{5'}$, $J_{4',5'} = 10.0$ Hz), 3.74 (dd, 1H, $H_{2'}$), 3.71 (dd, 1H, H_{3A} , $J_{2,3A} = 3.9$ Hz, $J_{3A,3B} = 10.9 \text{ Hz}$), 3.65 (t, 1H, $H_{4"}$, $J_{4",5"} = 9.4 \text{ Hz}$), 3.59 (dd, 1H, H_{3B} , $J_{2.3B} = 3.1 \text{ Hz}$), 3.58 (dd, 1H, $H_{2"}$), 3.52 (dd, 1H, $H_{4'}$), 3.46 (dd, 1H, $H_{6"A}$, $J_{5",6"A} = 2.9$ Hz, $J_{6"A,6"B} = 9.2$ Hz), 3.37 (dd, 1H, $H_{6"B}$, $J_{5".6"B}$ = 3.5 Hz), 2.40-2.27 (m, 6H, 3 x $CH_2\alpha$ myr.), 1.80-0.80 (m, 75H, CH₂/CH₃ myr.). ¹³C NMR (CDCl₃): δ 173.2, 173.0, 172.7 (C=O myr.), 138.6,

1,2-Di-O-palmitoyl-3-O-[2-O-(α -D-glucopyranosyl)-6-O-palmitoyl- α -D-glucopyranosyl]-sn-glycerol (1a). Fully protected glycolipid 29 (154 mg, 0.092 mmol) was dissolved in CH₂Cl₂/MeOH (10 mL, 1:4, v/v). Palladium on charcoal (10% Pd, 100 mg) was added and the heterogeneous mixture was hydrogenated at elevated pressure (3 atm.) in a Parr apparatus for 12 h. The catalyst was filtered off and the resulting filtrate was concentrated in vacuo and subjected to silica gel column chromatography (0-10% MeOH/CHCl₃) to afford glycolipid 1a as a white solid (100 mg, 0.088 mmol, 96%). $[\alpha]_D$ = +64.0 ° (c 0.2, MeOH). ¹H NMR (MeOD): δ 5.24 (m, 1H, H₂), 4.97 (d, 1H, H₁, J_{1',2'} = 3.3 Hz), 4.94 (d, 1H, $H_{1"}$, $J_{1",2"}$ = 3.7 Hz), 4.41 (dd, 1H, H_{1A} , $J_{1A,1B}$ = 12.1 Hz, $J_{1A,2}$ = 3.3 Hz), 4.35 (dd, 1H, $H_{6'A}$, $J_{5',6'A} = 2.4$ Hz, $J_{6'A,6'B} = 12.5$ Hz), 4.26 (dd, 1H, $H_{6'B}$, $J_{5',6'B} = 12.5$ Hz), 4.26 (dd, 1H, $H_{6'B}$, $J_{5',6'B} = 12.5$ Hz), 4.26 (dd, 1H, $H_{6'B}$, $J_{5',6'B} = 12.5$ Hz), 4.26 (dd, 1H, $H_{6'B}$), $J_{5',6'B} = 12.5$ Hz), 4.26 (dd, 1H, $H_{6'B}$), $J_{5',6'B} = 12.5$ Hz), 4.26 (dd, 1H, $H_{6'B}$), $J_{5',6'B} = 12.5$ Hz), 4.26 (dd, 1H, $H_{6'B}$), $J_{5',6'B} = 12.5$ Hz), 4.26 (dd, 1H, $H_{6'B}$), $J_{5',6'B} = 12.5$ Hz), 4.26 (dd, 1H, $H_{6'B}$), $J_{5',6'B} = 12.5$ Hz), 4.26 (dd, 1H, $H_{6'B}$), $J_{5',6'B} = 12.5$ Hz), 4.26 (dd, 1H, $H_{6'B}$), $J_{5',6'B} = 12.5$ Hz), 4.26 (dd, 1H, $H_{6'B}$), $J_{5',6'B} = 12.5$ Hz), 4.26 (dd, 1H, $H_{6'B}$), $J_{5',6'B} = 12.5$ Hz), 4.27 (dd, 1H, $H_{6'B}$), $J_{5',6'B} = 12.5$ Hz), 4.28 (dd, 1H, $H_{6'B}$), $J_{5',6'B} = 12.5$ Hz), 4.29 (dd, 1H, $H_{6'B}$), 4.29 (dd 5.4 Hz), 4.19 (dd, 1H, H_{1B} , $J_{1B,2} = 6.4$ Hz), 3.90 (dd, 1H, H_{3A} , $J_{2,3A} = 4.0$ Hz, $J_{3A,3B} = 4.0$ 10.8 Hz), 3.87 (m, 1H, $H_{5"}$), 3.78 (dd, 1H, $H_{3'}$, $J_{2',3'} = 8.9$ Hz, $J_{3',4'} = 9.4$ Hz), 3.72 (t, 1H, $H_{3"}$, $J_{2",3"} = J_{3",4"} = 9.1 \text{ Hz}$), 3.71-3.68 (m, 3H, $H_{5'}/H_{6"A}/H_{6"B}$), 3.62 (dd, 1H, $H_{2'}$), 3.60 (dd, 1H, H_{3B} , $J_{2.3B} = 3.0$ Hz), 3.46 (dd, 1H, $H_{4'}$, $J_{4'.5'} = 9.2$ Hz), 3.42 (dd, 1H, $H_{2"}$), 3.34 (dd, 1H, $H_{4''}$, $J_{4'',5''} = 8.6$ Hz), 2.40-2.28 (m, 6H, 3 x $CH_2\alpha$ palm.), 1.35-0.83 (m, 87H, CH₂/CH₃ palm.). ¹³C NMR (MeOD): δ 174.0, 173.9, 173.3 (C=O palm.), 96.3, 96.1 $(C_{1'}/C_{1''}),$ 75.9. 73.4, 72.0, 71.7, 71.6, 70.1, 69.8, $(C_2/C_2/C_3/C_4/C_5/C_2/C_3/C_4/C_5)$, 65.6 (C_3) , 63.2, 62.4, 61.5 $(C_1/C_6/C_6)$, 34.0-22.4 (CH₂ palm.), 13.8 (CH₃ palm.). MS (ESI): m/z 1132 (M+H)⁺.

Anal. Calcd for $C_{63}H_{118}O_{16}$ (1131.3): C, 66.87; H, 10.51. Found: C, 66.76; H, 10.60.

1,2-Di-*O*-myristoyl-3-*O*-[2-*O*-(α-D-glucopyranosyl)-6-*O*-myristoyl-α-D-glucopyranosyl]-sn-glycerol (1b). Fully protected glycolipid 30 (143 mg, 0.090 mmol) hydrogenated and purified as described for compound 1a to give glycolipid 1b as a white solid (88 mg, 0.085 mmol, 94%). [α]_D = +60.9 ° (*c* 0.2, MeOH). ¹H NMR (MeOD): δ 5.28 (m, 1H, H₂), 4.99 (d, 1H, H_{1'}, J_{1',2'} = 3.3 Hz), 4.94 (d, 1H, H_{1''}, J_{1'',2''} = 3.7 Hz), 4.40 (dd, 1H, H_{1A}, J_{1A,1B} = 12.1 Hz, J_{1A,2} = 3.5 Hz), 4.35 (dd, 1H, H_{6'A}, J_{5',6'A} = 2.6 Hz, J_{6'A,6'B} = 12.5 Hz), 4.29 (dd, 1H, H_{6'B}, J_{5',6'B} = 5.4 Hz), 4.19 (dd, 1H, H_{1B}, J_{1B,2} = 6.4 Hz), 3.91 (dd, 1H, H_{3A}, J_{2,3A} = 4.0 Hz, J_{3A,3B} = 11.0 Hz), 3.82 (m, 1H, H_{5''}), 3.76 (dd, 1H, H_{3''}, J_{2'',3''} = 8.9 Hz, J_{3',4''} = 9.4 Hz), 3.72 (t, 1H, H_{3''}, J_{2'',3''} = J_{3'',4''} = 9.1 Hz), 3.71-3.66 (m, 3H, H_{5'}/H_{6''A}/H_{6''B}), 3.62 (dd, 1H, H_{2'}), 3.60 (dd, 1H, H_{3B}, J_{2,3B} = 3.3 Hz), 3.40 (dd, 1H, H_{4'}, J_{4'',5''} = 9.2 Hz), 3.41 (dd, 1H, H_{2''}), 3.30 (dd, 1H, H_{4''}, J_{4'',5''} = 8.6 Hz), 2.43-2.19 (m, 6H, 3 x CH₂α myr.), 1.39-0.81 (m, 75H, CH₂/CH₃ myr.). ¹³C NMR (MeOD): δ 174.2, 173.9,

173.1 (C=O myr.), 96.4, 96.2 (C_1 / C_1 "), 76.1, 73.4, 72.0, 71.7, 71.6, 70.3, 69.8, 69.7, 69.5 (C_2 / C_2 / C_3 / C_4 / C_5 / C_2 "/ C_3 "/ C_4 "/ C_5 "), 66.8 (C_3), 63.3, 62.4, 61.5 (C_1 / C_6 "/ C_6 "), 35.1-22.9 (CH₂ myr.), 13.9 (CH₃ myr.). MS (ESI): m/z 1048 (M+H)⁺.

Anal. Calcd for $C_{57}H_{106}O_{16}$ (1047.1): C, 65.36; H, 10.20. Found: C, 65.30; H, 10.25.

ACKNOWLEDGEMENT

The work described in this paper was supported by the Netherlands Foundation for Chemical Research (SON) with financial aid from the Netherlands Organization for Scientific Research (NWO).

REFERENCES AND NOTES

- (a) W. Curatolo, Biochim. Biophys. Acta, 906, 111 (1987); (b) W. Curatolo, Biochim. Biophys. Acta, 906, 137 (1987); (c) R.D. Koynova, B.G. Tenchov, H. Kuttenreich and H.J. Hinz, Biochemistry, 32, 12437 (1993).
- C.A.A. van Boeckel and J.H. van Boom, Tetrahedron, 41, 4545 (1985).
- (a) W. Fischer and H. Landgraf, Biochim. Biophys. Acta, 380, 227 (1975); (b) H.U. Koch and W. Fischer, Biochemistry, 17, 5275 (1978); (c) W. Fischer, D. Schuster and R.A. Laine, Biochim. Biophys. Acta, 575, 389 (1979).
- 4. K. Fukase, T. Yoshimura, S. Kotani and S. Kusumoto, Bull. Chem. Soc. Jpn., 67, 473 (1994).
- 5. Prof. dr. S. Arai, personal communication.
- 6. R.L. Halcomb and S.J. Danishefsky, J. Am. Chem. Soc., 111, 6661 (1989).
- 7. D.M. Gordon and S.J. Danishefsky, *Carbohydr. Res.*, **206**, 361 (1990).
- 8. N.J. Davies and S.L. Flitsch, Tetrahedron Lett., 34, 1181 (1993).
- 9. A.G.M. Barett and A.C. Lee, J. Org. Chem., 57, 2818 (1992).
- Recent examples of the application of anomeric phosphates as glycosyl donors: (a)
 S. Hashimoto, T. Honda and S. Ikegami, J. Chem. Soc., Chem. Commun., 685 (1989); (b) H.I. Duynstee, E.R. Wijsman, G.A. van der Marel and J.H. van Boom, Synlett, 313 (1996).
- 11. L.A.J.M. Sliedregt, Thesis, Leiden (1994).
- M.S. Shakhani, K.M. Khan, K. Mahmood, P.M. Shah and S. Malik, *Tetrahedron Lett.*, 31, 1669 (1990).
- 13. F. Weygand and H. Zeimann, *Liebigs Ann. Chem.*, **657**, 179 (1962).
- 14. G.H. Veeneman, S.H. van Leeuwen, H.M. Zuurmond and J.H. van Boom, J. Carbohydr. Chem., 9, 783 (1990).
- (a) T. Ogawa, H. Beppu and S. Nakabayashi, Carbohydr. Res., 93, c6 (1981); (b) H. Paulsen and M. Paal, Carbohydr. Res., 135, 53 (1984).
- 16. P.A. Gent and R. Gigg, Chem. Phys. Lipids, 16, 111 (1976).